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(FILE 'HOME' ENTERED AT 09:10:03 ON 22 SEP 2004)

FILE 'MEDLINE, CAPLUS' ENTERED AT 09:11:00 ON 22 SEP 2004

FILE 'MEDLINE, BIOSIS, CAPLUS' ENTERED AT 09:11:11 ON 22 SEP 2004

L1 18 S (RAPID?) (9A) (AMPLIF?) (9A) BIOPS?
L2 8 DUP REM L1 (10 DUPLICATES REMOVED)
L3 1642 S RAPID? AND (AMPLIF? OR PCR OR POLYMERASE (W)CHAIN) AND BIOPS?
L4 1642 S RAPID? AND (AMPLIF? OR PCR OR POLYMERASE (W)CHAIN) AND BIOPS?
L5 271 S L4 AND PY<1994
L6 138 DUP REM L5 (133 DUPLICATES REMOVED)

FILE 'STNGUIDE' ENTERED AT 09:21:31 ON 22 SEP 2004

FILE 'MEDLINE, BIOSIS, CAPLUS' ENTERED AT 09:22:36 ON 22 SEP 2004

FILE 'STNGUIDE' ENTERED AT 09:22:36 ON 22 SEP 2004

FILE 'MEDLINE, BIOSIS, CAPLUS' ENTERED AT 09:23:25 ON 22 SEP 2004

L7 325 S PHENOL (9A) (PCR OR POLYMERASE (W)CHAIN)
L8 45 S L7 AND PY<1994
L9 27 DUP REM L8 (18 DUPLICATES REMOVED)

FILE 'STNGUIDE' ENTERED AT 09:28:18 ON 22 SEP 2004

FILE 'MEDLINE, BIOSIS, CAPLUS' ENTERED AT 09:32:05 ON 22 SEP 2004

L10 53 S "WITHOUT" (9A) (ETHANOL OR ETOH) (9A) (PRECIP? OR PPT)
L11 37 DUP REM L10 (16 DUPLICATES REMOVED)

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L9 ANSWER 23 OF 27 CAPLUS COPYRIGHT 2004 ACS on STN
AN 1991:1750 CAPLUS
DN 114:1750
TI DNA isolated from plastic embedded tissue is suitable for PCR
AU Gruenewald, K.; Feichtinger, H.; Weyrer, K.; Dietze, O.; Lyons, J.
CS Dep. Intern. Med., Univ. Innsbruck, Austria
SO Nucleic Acids Research (1990), 18(20), 6151
CODEN: NARHAD; ISSN: 0305-1048
DT Journal
LA English
AB Plastic embedding of routinely formalin fixed tissues preserves excellent morphol. and is used in many centers especially for routine histol. of bone marrow biopsies. Since recent reports have shown the feasibility of polymerase chain reaction (PCR) amplification of DNA isolated from fixed paraffin embedded tissue, DNA was extracted from bone marrow trephine biopsies fixed in 10% formalin, decalcified in EDTA, embedded in Technovit 700 (hydroxyethylmethacrylate; HEMA; Kulzer, FRG) and stored between 2 mo and 4 yr at room temperature to test also for the influence of different ds.p. of the embedding medium. This report describes 2 alternate methods used to extract the DNA from plastic embedded tissue for subsequent amplification by PCR. One method uses proteinase K digestion and extns. with **phenol**/chloroform/isoamyl alc. prior to **PCR** while the 2nd involves direct amplification of proteinase K digested supernatants. Length of proteinase K digestion and temperature were found to influence DNA recovery dramatically. Different storage times i.e. different ds.p. of the embedding medium HEMA, had no influence on DNA yield or degradation DNA isolated by either method 1 or 2 was suitable for amplification by PCR. The second method was used to amplify a 526 bp fragment of a Gs protein α chain gene.

L9 ANSWER 9 OF 27 MEDLINE on STN DUPLICATE 6
 AN 93216961 MEDLINE
 DN PubMed ID: 8385159
 TI Simple technique for detecting RNA viruses by PCR in single sections of wax embedded tissue.
 AU Woodall C J; Watt N J; Clements G B
 CS Regional Virus Laboratory, Ruchill Hospital, Glasgow.
 SO Journal of clinical pathology, (1993 Mar) 46 (3) 276-7.
 Journal code: 0376601. ISSN: 0021-9746.
 CY ENGLAND: United Kingdom
 DT Journal; Article; (JOURNAL ARTICLE)
 LA English
 FS Abridged Index Medicus Journals; Priority Journals
 EM 199305
 ED Entered STN: 19930521
 Last Updated on STN: 19970203
 Entered Medline: 19930506
 AB The detection of specific RNA species in wax-embedded tissue sections using the polymerase chain reaction (PCR) means that gene expression can be studied and RNA viruses detected in stored histological tissue samples. This technique potentially allows the distribution of gene expression and viral replication to be studied in finely subdivided tissues. A technique is presented that has been used successfully to detect short RNA target sequences (130-420 bases) from proto-oncogene Abelson, human enteroviruses, and the sheep retrovirus Maedi-Visna virus using RNA PCR in single wax sections (20-30 microns). Various tissues were used which had not been deliberately prepared for this purpose. In a simple procedure hot xylene dewaxing is followed by acid **phenol** extraction of RNA and RNA **PCR**.

L6 ANSWER 22 OF 138 MEDLINE on STN DUPLICATE 15
 AN 93358930 MEDLINE
 DN PubMed ID: 8354305
 TI Detection of Helicobacter pylori in gastric **biopsy** tissue by
polymerase chain reaction.
 AU Wang J T; Lin J T; Sheu J C; Yang J C; Chen D S; Wang T H
 CS Department of Internal Medicine, National Taiwan University Hospital,
 Taipei.
 SO European journal of clinical microbiology & infectious diseases : official
 publication of the European Society of Clinical Microbiology, (1993
May) 12 (5) 367-71.
 Journal code: 8804297. ISSN: 0934-9723.
 CY GERMANY: Germany, Federal Republic of
 DT Journal; Article; (JOURNAL ARTICLE)
 LA English
 FS Priority Journals
 EM 199309
 ED Entered STN: 19931008
 Last Updated on STN: 19931008
 Entered Medline: 19930921
 AB To evaluate the sensitivity of a **polymerase chain**
 reaction (**PCR**) assay using nested primers in detecting
 Helicobacter pylori, gastric tissue **biopsy** specimens were
 collected on endoscopy from 17 patients with a duodenal ulcer. DNA was
 extracted by phenol/chloroform treatment or boiling in water, and then
 subjected to a nested **PCR** using two primer pairs from the urease
 gene of Helicobacter pylori. Fourteen of the 17 patients were positive
 for Helicobacter pylori using DNA samples extracted by either method. The
PCR results correlated well with the results of an enzyme
 immunoassay to detect IgG antibody. However, there were two culture
 negative patients. The three **PCR** negative patients were both
 culture negative and serologically negative. DNA from 9 of the 14
 patients was randomly selected and subjected to semiquantification by
 serial dilutions, and then **PCR**. The results showed that
 phenol/chloroform extraction yielded 10-1000 times more DNA than the
 boiling method. It is concluded that the **PCR** assay is a
rapid and sensitive method for detecting Helicobacter pylori, and
 that phenol/chloroform extraction is superior to simple boiling in
 obtaining DNA samples for **PCR**.

L6 ANSWER 70 OF 138 MEDLINE on STN DUPLICATE 48
 AN 92202313 MEDLINE
 DN PubMed ID: 1313040
 TI A simple and **rapid** method of high quantity DNA isolation from
 cervical scrapes for detection of human papillomavirus infection.
 AU Gopalkrishna V; Francis A; Sharma J K; Das B C
 CS Division of Molecular Oncology, Institute of Cytology and Preventive
 Oncology (ICMR), New Delhi, India.
 SO Journal of virological methods, (1992 Jan) 36 (1) 63-72.
 Journal code: 8005839. ISSN: 0166-0934.
 CY Netherlands
 DT Journal; Article; (JOURNAL ARTICLE)
 LA English
 FS Priority Journals
 EM 199204
 ED Entered STN: 19920509
 Last Updated on STN: 19920509
 Entered Medline: 19920428
 AB Infection with human papillomavirus (HPV) is an important etiological
 factor in the development of cervical cancer, and detection of the viral
 genome is of prognostic importance, particularly for preneoplastic
 lesions. We developed a simple, easy and efficient non-organic method of
 DNA extraction from cervical scrapes for reliable detection of HPV DNA
 sequences. The method involves incubation of cell nuclei in higher
 concentration of proteinase K at 65 degrees C for 2.5 h. Following
 prolonged incubation at higher temperature, the enzyme is autoinactivated
 and the DNA isolated can be used directly for analysis without further
 purification. The recovery of DNA is more than 95% and it can be easily
 cleaved by restriction enzymes and is suitable for **amplification**
 by the **polymerase chain** reaction (PCR). The
 whole procedure is carried out in a single Eppendorf tube and a large
 number of specimens can be processed at a time without any error of
 handling. DNA extracted from a single smear sample is sufficient to
 conduct as many as four different molecular biology tests. This provides
 an opportunity for verification of sensitivity, specificity and
 reliability of each test for diagnosis of HPV infection without resorting
 to **biopsy**.

L6 ANSWER 124 OF 138 MEDLINE on STN DUPLICATE 81
AN 89356258 MEDLINE
DN PubMed ID: 2548820
TI A **rapid** procedure to identify newborn transgenic mice.
AU Lin C S; Magnuson T; Samols D
CS Department of Genetics, School of Medicine, Case Western Reserve
University, Cleveland, OH 44106.
NC AR20618 (NIAMS)
P30CA43703 (NCI)
SO DNA (Mary Ann Liebert, Inc.), (1989 May) 8 (4) 297-9.
Journal code: 8302432. ISSN: 0198-0238.
CY United States
DT Journal; Article; (JOURNAL ARTICLE)
LA English
FS Priority Journals
EM 198910
ED Entered STN: 19900309
Last Updated on STN: 19980206
Entered Medline: 19891006
AB We have developed a **rapid** procedure to identify newborn
transgenic mice containing foreign genetic material in their genome. The
protocol involves collagenase digestion of a small amount of tail tissue
which can be taken very early after birth, phenol and chloroform
extraction, **polymerase chain** reaction, and
polyacrylamide gel electrophoresis. The entire procedure, from tissue
biopsy to final results, can be completed in 1 day.

AN 90104461 MEDLINE
DN PubMed ID: 2557850
TI Method of extracting DNA from fine needle aspirates of human solid tumors for Southern blot analysis.
AU Harnett P R; Greenberg M L; Tattersall M H; Kefford R F
CS Medical Oncology Unit, Institute of Clinical Pathology, Westmead, New South Wales, Australia.
SO Analytical and quantitative cytology and histology / the International Academy of Cytology [and] American Society of Cytology, (1989 Dec) 11 (6) 375-8.
Journal code: 8506819. ISSN: 0884-6812.
CY United States
DT Journal; Article; (JOURNAL ARTICLE)
LA English
FS Priority Journals
EM 199002
ED Entered STN: 19900328
Last Updated on STN: 19900328
Entered Medline: 19900222
AB A **rapid**, simple, convenient method for extracting DNA from fine needle aspiration (FNA) samples of human solid tumors for Southern blot hybridization studies is described. After the preparation of an air-dried cytologic smear, the remaining sample in the needle was rinsed directly into a test tube for DNA extraction. The extraction procedure, in which manipulation of the sample is minimized, produced sufficient DNA for Southern blot analysis within 24 hours of the FNA **biopsy** in the ten consecutive cases studied. The DNA bound to the nylon membranes can be washed and reexamined with a variety of probes, allowing studies of lymphoid cell lineage, oncogene **amplification** or tumor progression. The assessment of cellularity on the cytologic specimen at the time of FNA provided a reliable guide to the need for further passes to obtain sufficient cells for DNA hybridization; the cytologic diagnosis could also be made on the smears.

ANSWER 14 OF 27 MEDLINE on STN

AN 93187424 MEDLINE
DN PubMed ID: 1293226
TI Comparison of polymerase chain reaction and IDEIA Chlamydia in detection of Chlamydia trachomatis from first-voided urine of male urethritis patients.
AU Komeda H; Deguchi T; Tada K; Yamamoto H; Kanematsu E; Iwata H; Okano M; Ito Y; Saito A; Ban Y; +
CS Department of Urology, Gifu University School of Medicine.
SO Kansenshogaku zasshi. Journal of the Japanese Association for Infectious Diseases, (1992 Oct) 66 (10) 1473-8.
Journal code: 0236671. ISSN: 0387-5911.
CY Japan
DT Journal; Article; (JOURNAL ARTICLE)
LA Japanese
FS ~~Priority Journals~~
EM 199304
ED Entered STN: 19930416
Last Updated on STN: 19930416
Entered Medline: 19930402
AB We have reported a method for detection of Chlamydia trachomatis by polymerase chain reaction (PCR) with two oligonucleotides based on sequences within the major outer membrane protein gene from C. trachomatis serovar L2. In the previous report, in addition to treatment of the mixture of first-voided urine (FVU) sediment and 1 ml of urine with proteinase K. DNA purification by **phenol** extraction was necessary for preparation of template DNA for PCR. In this study, FVU sediment was suspended in 1 ml of Chlamydiazyme dilution buffer and a part of the suspension was treated with proteinase K for DNA extraction. The DNA extraction solution could be used as template for PCR without purification of DNA by **phenol** extraction. One hundred FVU specimens obtained from male urethritis patients were examined with the two methods (PCR and IDEIA) for detection of C. trachomatis. In 33 of 100 specimens, the DNA fragments of C. trachomatis was amplified by the PCR and in 32 of 100, the chlamydial antigen was detected by IDEIA. The positive and negative coincidence rate of the PCR to IDEIA were 93.8% (30/32) and 95.6% (65/68) respectively, resulting in a high overall coincidence rate at 95%. Thus, the improved method with PCR using FVU as a specimen is proved to be a useful, non-invasive diagnostic tool for diagnosis of chlamydial urethritis.